

Bio-Control Efficiency of Water Bug *Diplonychus Rusticus* (Hemiptera: Belostomatidae) on *Culex*mosquito Larvae

OPEN ACCESS

Volume: 12

Special Issue: 1

Month: October

Year: 2024

E-ISSN: 2582-0397

P-ISSN: 2321-788X

Impact Factor: 3.025

Citation:

Abhilash, HR, et al. "Bio-Control Efficiency of Water Bug *Diplonychus Rusticus* (Hemiptera: Belostomatidae) on *Culex*mosquito Larvae." *Shanlax International Journal of Arts, Science and Humanities*, vol. 12, no. S1, 2024, pp. 71–76.

DOI:

<https://doi.org/10.34293/sijash.v12iS1-i2-Oct.8421>

H.R. Abhilash

*Department of Zoology, Yuvaraja's College (Autonomous)
University of Mysore, Mysuru, Karnataka, India*

K.R. Shashank

*Department of Zoology, Yuvaraja's College (Autonomous)
University of Mysore, Mysuru, Karnataka, India*

M. Mahadevaswamy

*Department of Zoology, Yuvaraja's College (Autonomous)
University of Mysore, Mysuru, Karnataka, India*

S.V. Sathish

*Associate Professor of Zoology, Department of Zoology
Sri Mahadeshwara Government First Grade College
Kollegal, Chamarajanagara (Dist.), Karnataka, India*

Abstract

*Mosquito-borne diseases continue to pose a significant public health risk, especially in tropical regions, due to the widespread occurrence of malaria, dengue, chikungunya, and other arboviruses. The extensive use of chemical insecticides has led to resistance in mosquito populations and has been proven to be harmful to a host of non-target organisms. As an environmentally friendly alternative, biological control using natural predators has gained attention. This study evaluates the bio-control efficiency of the water bug *Diplonychus rusticus* (Hemiptera: Belostomatidae) against *Culex* mosquito larvae, a known vector of many diseases. Specimens of *D. rusticus* were collected from Dalvoy Lake, Mysore, and tested in controlled laboratory conditions for their predatory impact (PI) and clearance rate (CR) on fourth instar *Culex* larvae. The results showed that *D. rusticus* effectively preys on *Culex* larvae, with a predatory impact of 3.13 larvae per hour and a clearance rate of 2.167 larvae per liter/day. The findings suggest that *D. rusticus* holds potential as a bio-control agent, offering a promising alternative to chemical interventions in vector control programs. Further research into the application of this predator in natural aquatic habitats could aid in managing mosquito populations and mitigating the widespread dissemination of mosquito-borne diseases.*

Keywords: Mosquito-Borne Diseases, *Diplonychus Rusticus*, *Culex* Larvae, Clearancerate

Introduction

Mosquito-borne diseases like dengue, chikungunya, lymphatic filariasis, yellow fever, Japanese encephalitis, malaria, and a range of other arboviruses remain serious public health concerns in many tropical nations (Eba et al.). There has been increased fear about the growing prevalence and geographical expansion of mosquito-borne

diseases worldwide, with new places witnessing the onset of these diseases (Franklinos et al.). The use of chemicals to control mosquito populations has negative consequences for non-target species as well as creating chemically resistant variants (WHO). This has led to a greater emphasis on sustainable alternative methods of managing vector populations by biological means, for instance, making use of natural predators of mosquito immatures (Karunaratne and Hemingway; Mandal et al.). A wide range of living organisms have been identified as possible mosquito control agents, including bacteria, protozoa, fungus, nematodes, other invertebrate and vertebrate predators (Aditya et al.; Chandra et al.; Gautam et al.; Lundkvist et al.; Stav et al.; Venkatesh and Tyagi).

Of the various predators mentioned above, predatory aquatic insects hold great potential as biological control agents, especially the members of the order Hemiptera, Odonata, Coleoptera, and Diptera (Lutziatigripes), which are cosmopolitans and locally available (Saha et al.; Shaalan and Canyon). The water bugs in the family Belostomatidae (Heteroptera) are well recognized predators of aquatic snails and insects, including several immature stages of mosquitoes (Brahma et al.; Ohba and Nakasuji). Several workers believe that their predatory nature makes them suitable for biological control of mosquito larvae (Ghosh and Chandra; Kumar and Hwang; Weterings et al.). Thus, the purpose of this study was to evaluate the predatory efficiency of hemipteran bug *Diplonchus rusticus* using the larvae of *Culex* mosquito as prey. *D. rusticus*, generally referred to as a water bug, is a common inhabitant and a major predator in the aquatic ecosystems of Mysore and its surrounding areas (Abhilash et al.). The findings will serve as the basis for evaluating these predators as biological resources against *Culex* sp. and other mosquito vectors.

Materials and Methods

The adult morphs of *D. rusticus* were collected from Dalvoy Lake of Mysore district, Karnataka, India, by dragging a circular pond net (500-µm mesh size) through the vegetation of the littoral zone. We brought the water bugs to the laboratory, transferred them to plastic trays containing 2 L of lake water, and fed them ad libitum mosquito larvae (*Culex* sp.) as food. They were kept in the laboratory setup for 5 days for acclimatization before starting the experiment. Mosquito larvae were collected from the sewage drains near Dalvoy Lake for feeding the water bugs during the acclimation period. Further, the egg rafts of *Culex* mosquito were collected from the same site and were placed within an enamel tray of 30 x 25 cm capacity containing tap water. Upon hatching, the larvae were given a powdered mixture of yeast granules and fish food. The following experiments were conducted to evaluate the rate of predation of the water bug on *Culex* sp. larvae.

Determination of Predatory Impact (PI)

In this experiment, the predatory impact was determined by following the method adopted by Jacob et al., where a single predatory insect was supplied with 25 *Culex* larvae in a 500ml beaker (in the ratio 1:25) containing 400 ml of water. The experiment included three replicates along with a control group containing only *Culex* larvae. Observations were made over a total period of 8 hours. At the conclusion of each hour, the number of preys consumed and those killed in the control group was documented. After each hour, new mosquito larvae were introduced to replace those that had been eaten or killed. The predatory impact was calculated by following the method adopted by Saha et al., using the formula:

$$PI = \left(\sum PE \right) / T$$

Where, PI is the Predatory impact (No. of prey consumed/hours); PE = No. of prey eaten or killed; T=Time (here, T = 8 hours).

Determination of Clearance Rate (CR)

In this experiment the clearance rate (CR) was determined by offering 2 predators with 100 IV instars Culexlarvae within a circular plastic tub containing 1litre pond water. The count of larvae killed or consumed was recorded, and the prey density was reset every 24 hours for three consecutive days, using the same group of predators. Three replicates plus a control group with only Culexlarvae were set for the experiment. The data collected from the experiment were used to calculate clearance rate (CR) using the method established by Gilbert & Burns, as adopted by Aditya & Saha.

$$CR = \frac{V (\ln P)}{TN}$$

Where, CR = Clearance rate of predators (Nos. of prey killed/liters/day/predator); V is the Volume of water (litr); T is the Time (in day), N is the Number of predators, and P is the Nos. of prey killed.

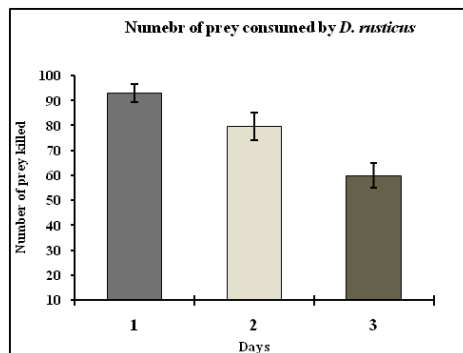


Figure 1 Mean Number (Mean ± SE) of IV Instar Larvae Killed per day by Two Adult D Rusticus (for three consecutive days)

Table 1 Number of Larvae Killed/Hour by D. Rusticus

Time (hour)	No. of larva Killed/Eaten (Mean of 3 replicates)
1	5.33
2	2.67
3	0.67
4	4.67
5	6.00
6	2.67
7	0.67
8	2.33
Total	25.00
PI	25/8 = 3.13

Table 2 Clearance Rate of D. Rusticus on IV Instar Culex Larva

	Vol (l)	No. Killed/day	ln	TN	CR
Day 1	1	93	4.53	2	2.266
Day 2	1	79.67	4.38	2	2.189
Day 3	1	60	4.09	2	2.047
				Mean	2.167

*Where, CR = Clearance rate of predators (Nos. of prey killed /liters/day/predator); V is the Volume of water (litr); T is Time in days (1 day), N is Number of predators (2), ln = natural logarithm; P = Nos. of prey killed.

Result and Discussion

In the present experiment, the predatory efficiency of the water bug, *D. rusticus*, was assessed using *Culex* mosquito larvae. It was observed that the water bug vigorously seized the mosquito larva with its pro- and mesothoracic legs, immobilized it by piercing with its sharp rostrum, and then consumed its internal body fluids. It was noted that a large number of preys were simply killed without sucking the body fluids. A previous study also reported similar findings, indicating that *D. rusticus* killed more mosquito larvae than it actually fed (Saha et al.). The predatory impact demonstrates the predator's killing efficiency over a specific time period. In the present experiment PI values for *D. rusticus* for 4th instar *Culex* larvae was found to be 3.13 larvae/hour at prey density of 25 larvae (Table 1). The rate of prey consumption is influenced by the size and density of prey species, with many arthropodan predators exhibiting a preference for larger prey, specifically fourth and fifth instar larvae, over smaller alternatives (Gurumoorthy et al.; Prabakaran). The clearance rate indicates the combined effects of the predator's search efficiency, predation, and consumption, along with the evasive strategies of the prey, within a defined time and area (Gautam et al.). Collectively two adults of *D. rusticus* consumed between 60 to 93 fourth-instar *Culex* larvae per day (Fig. 1). The clearance rate (CR) was found to be 2.167 prey larvae litres/day/predator (Table 2). The number of prey consumed varied drastically across the days, indicating a decrease in the predation rate after reaching maximum satiation, which is a typical predation pattern for *D. rusticus* (Pramanik and Raut).

The results of this study on the predatory efficacy of *D. rusticus* against *Culex* mosquito larvae have significance for developing bio-control methods as part of an integrated vector control program in India. Given the challenges in controlling adult mosquitoes due to extensive insecticide resistance, using potential aquatic insect predators may serve as an alternate or complementary strategy for controlling the adult mosquito population by targeting the immature stages of *Culex* mosquitoes. The outcome of the current laboratory experiment suggests a comparable potential for utilizing these hemipteran bugs as predators of mosquito larvae in various aquatic habitats in Mysore and its surrounding regions.

Acknowledgment

The authors are grateful to The Principal, Yuvaraja's College (Autonomous), University of Mysore, Mysore, India for the facilities provided.

References

1. Abhilash, H. R., et al. 'Biodiversity and Abundance of Aquatic Insects in Two Freshwater Lakes of Mysore District, Karnataka, India'. *Arthropods*, vol. 12, no. 1, 2023, pp. 1–15.
2. Aditya, G, et al. 'Frequency-Dependent Prey-Selection of Predacious Water Bugs on *Armigeres Subalbatus* Immatures'. *Journal of Vector Borne Diseases*, vol. 42, no. 1, 2005, pp. 9–14.
3. Aditya, Gautam, et al. Predatory Activity of *Rhantus Sikkimensis* and Larvae of *Toxorhynchites Splendens* on Mosquito Larvae in Darjeeling, India. 2006, pp. 66–72.
4. Aditya, Gautam, and Goutam Kumar Saha. 'Predation of the Beetle *Rhantus Sikkimensis* (Coleoptera: Dytiscidae) on the Larvae of *Chironomus Meigen* (Diptera: Chironomidae) of the Darjeeling Himalayas of India'. *Limnologica*, vol. 36, no. 4, 2006, pp. 251–57. Science Direct, <https://doi.org/10.1016/j.limno.2006.07.004>.

5. Brahma, S., et al. 'Influence of Density on Intraguild Predation of Aquatic Hemiptera (Heteroptera): Implications in Biological Control of Mosquito'. *Journal of Entomological and Acarological Research*, vol. 46, no. 1, 2014, pp. 6–12.
6. Chandra, G., et al. 'Mosquito Control by Larvivorous Fish'. *Indian Journal of Medical Research*, vol. 127, no. 1, 2008, p. 13.
7. Eba, Kasahun, et al. 'Bio-Control of Anopheles Mosquito Larvae Using Invertebrate Predators to Support Human Health Programs in Ethiopia'. *International Journal of Environmental Research and Public Health*, vol. 18, no. 4, 4, 2021, p. 1810. www.mdpi.com, <https://doi.org/10.3390/ijerph18041810>.
8. Franklino, Lydia H. V., et al. 'The Effect of Global Change on Mosquito-Borne Disease'. *The Lancet Infectious Diseases*, vol. 19, no. 9, Sept. 2019, pp. e302–12. www.thelancet.com, [https://doi.org/10.1016/S1473-3099\(19\)30161-6](https://doi.org/10.1016/S1473-3099(19)30161-6).
9. Ghosh, Anupam, and Goutam Chandra. 'Functional Responses of *Laccotrephes Griseus* (Hemiptera: Nepidae) against *Culex Quinquefasciatus* (Diptera: Culicidae) in Laboratory Bioassay'. *J Vector Borne Dis*, vol. 48, no. 2, 2011, p. 72.
10. Gilbert, John J., and Carolyn W. Burns. 'Some Observations on the Diet of the Backswimmer, *Anisops Wakefieldi* (Hemiptera: Notonectidae)'. *Hydrobiologia*, vol. 412, no. 0, 1999, pp. 111–18. Springer Link, <https://doi.org/10.1023/A:1003812718853>.
11. Gurumoorthy, K., et al. 'Predatory Behaviour and Efficiency of the Water Bug *Sphaerodema rusticum* on Mosquito Larvae *Culex quinquefasciatus*.' *International Journal of Pure and Applied Zoology*, vol. 1, no. 1, 2013, pp. 24–29.
12. Jacob, Sonia, et al. 'Bio Control Efficiency of Odonata Nymphs on *Aedes Aegypti* Larvae'. *IOSR J. Environ. Sci. Toxicol. Food Technol*, vol. 11, no. 9, 2017, pp. 1–4.
13. Karunaratne, S. H. P. P., and J. Hemingway. 'Insecticide Resistance Spectra and Resistance Mechanisms in Populations of Japanese Encephalitis Vector Mosquitoes, *Culex Tritaeniorhynchus* and *Cx. Gelidus*, in Sri Lanka'. *Medical and Veterinary Entomology*, vol. 14, no. 4, 2000, pp. 430–36. Wiley Online Library, <https://doi.org/10.1046/j.1365-2915.2000.00252.x>.
14. Kumar, Ram, and Jiang-Shiou Hwang. 'Larvicidal Efficiency of Aquatic Predators: A Perspective for Mosquito Biocontrol'. *Zoological Studies*, 2006.
15. Lundkvist, E., et al. 'Diving Beetles (Dytiscidae) as Predators of Mosquito Larvae (Culicidae) in Field Experiments and in Laboratory Tests of Prey Preference'. *Bulletin of Entomological Research*, vol. 93, no. 3, June 2003, pp. 219–26. Cambridge University Press, <https://doi.org/10.1079/BER2003237>.
16. Mandal, S. K., et al. 'Biocontrol Efficiency of Odonate Nymphs against Larvae of the Mosquito, *Culex Quinquefasciatus* Say, 1823'. *Acta Tropica*, vol. 106, no. 2, 2008, pp. 109–14. Science Direct, <https://doi.org/10.1016/j.actatropica.2008.02.002>.
17. Ohba, Shin-ya, and Fusao Nakasuji. 'Density-Mediated Indirect Effects of a Common Prey Tadpole on Interaction between Two Predatory Bugs: *Kirkaldyia Deyrolli* and *Laccotrephes Japonensis*'. *Population Ecology*, vol. 49, no. 4, 2007, pp. 331–36. Springer Link, <https://doi.org/10.1007/s10144-007-0051-7>.
18. Prabakaran, V. 'Predatory Potential of *Diplonychus Indicus* (Hemiptera: Belostomatidae) on Mosquito Larva'. *Journal of Ecobiology*, vol. 4, 1992, pp. 169–75.
19. Pramanik, M. K., and S. K. Raut. 'Predation Rhythm in the Water-Bug *Sphaerodema Rusticum Fabricius*'. *Environ Ecol*, vol. 23, 2005, pp. 707–13.
20. Saha, Nabaneeta, et al. 'A Comparative Study of Predation of Three Aquatic Heteropteran Bugs on *Culex Quinquefasciatus* Larvae'. *Limnology*, vol. 8, no. 1, 2007, pp. 73–80. Springer Link, <https://doi.org/10.1007/s10201-006-0197-6>.

21. Shaalan, Essam Abdel-Salam, and Deon V. Canyon. 'Review Paper Aquatic Insect Predators and Mosquito Control'. *Tropical Biomedicine*, no. 26, 2009, pp. 223–61.
22. Stav, Gil, et al. 'Individual and Interactive Effects of a Predator and Controphic Species on Mosquito Populations'. *Ecological Applications*, vol. 15, no. 2, 2005, pp. 587–98. Wiley Online Library, <https://doi.org/10.1890/03-5191>.
23. Venkatesh, A., and B. K. Tyagi. 'Predatory Potential of *Bradinopyga Geminata* and *Ceriagrion Coromandelianum* Larvae ON Dengue Vector *Aedes Aegypti* under Controlled Conditions (Anisoptera: Libellulidae; Zygoptera: Coenagrionidae; Diptera: Culicidae)'. *Odonatologica*, vol. 42, no. 2, 2013, pp. 139–49.
24. Weterings, R., et al. 'Factors Influencing the Predation Rates of *Anisops Breddini* (Hemiptera: Notonectidae) Feeding on Mosquito Larvae'. *Journal of Entomological and Acarological Research*, vol. 46, no. 3, 2014, p. 107. DOI.org (Crossref), <https://doi.org/10.4081/jear.2014.4036>.
25. WHO. WHO [World Health Organization]. *Manual on Practical Entomology in Malaria. Part II. Methods and Techniques*. WHO Offset Publ. 13., 1975.